

in simple dialkylammonium membrane ($2C_nN^+2C_1$, $n = 14, 16, 18$) was in the reversed direction of that mentioned above; e.g., for $2C_{16}N^+2C_1$ membrane, $\lambda_{\max} = 388$ nm in the low temperature range (20 - 25°C) and 408 nm in the high temperature range (above 29°C) (Fig. 1b). The spectral shift occurs at 26 - 28°C in accord with T_c of this membrane (28°C). Similar T_c dependence was found for the membrane matrix in which the glutamic acid moiety in $\underline{1}$ is replaced by the L-aspartic acid moiety.

In the case of $\underline{1}$ ($n = 4$), the identical spectral change was observed regardless of whether $\underline{1}$ contains L-, D-, or DL-glutamic acid residue. This indicates that the chiral property of the membrane is not directly responsible for the spectral change. On the other hand, the length of the "spacer" methylene chain, n in $\underline{1}$, is crucial for the spectral shift. The same spectral change as mentioned above for L- $\underline{1}$ ($n = 4$) membrane is found in the case of $n = 3$; however, when $n = 2$, λ_{\max} is located at 367 nm at temperature below T_c (27°C) and shifts to 415 nm upon heating to temperature above T_c . With $n = 5$ and 10, the T_c dependence is not observed: λ_{\max} , 385 to 405 nm. λ_{\max} for L- $\underline{1}$ ($n = 6$) is at 440 nm at temperature below T_c (42°C) and at 424 nm above T_c . The spectral shift is only 16 nm. These data indicate that the change in the spacer length by one methylene unit causes large differences in the spectral property.

Representative temperature dependence of λ_{\max} is shown in Fig. 2. Sharp changes in λ_{\max} are observed at T_c in the membrane matrix of $\underline{1}$ ($n = 2$ and 4) in the opposite directions, but the change is less drastic for $2C_{16}N^+2C_1$ membrane.

The observed λ_{\max} values cover a range of 367 to 488 nm, as against the "normal" λ_{\max} value of 420 - 430 nm in organic media. The blue shift beyond the normal λ_{\max} range is caused by stacking of the dye molecules in the parallel orientation (H-type aggregation), as already discussed in the micellar and other systems.²⁻⁴ On the other hand, the red shift may be attributed to dye stacking in the head-to-tail orientation (J-like aggregation).⁹⁻¹⁰

The influence of dye aggregation on the λ_{\max} shift is clearly shown in Fig. 3. The largest red shift is observed in the rigid membrane matrix of L- $\underline{1}$ ($n = 4$) at the molar ratio of 1:10 ([Methyl Orange]/[L- $\underline{1}$ ($n = 4$)]). The red shift decreases with dilution of Methyl Orange and λ_{\max} reaches 455 nm at the ratio of 1:500. It is presumed that the aggregated dye is not present at the latter molar ratio, and the dye stacking alone should account for the red shift of 33 nm (488 - 455 nm). The λ_{\max} value is independent of the molar ratio in the fluid membrane matrix at temperature above T_c . This λ_{\max} value (424 nm) is typical of that in common organic media, indicating that dye aggregation and/or specific interaction with the membrane is not significant in the case of the fluid membrane. The λ_{\max} difference of 455 - 424 = 31 nm at the molar ratio of 1/100 to 1/500 is thus produced by the change in the microenvironment of the membrane due to phase transition. Since the λ_{\max} value in the fluid matrix is normal, the 31-nm red shift may be ascribed to the specific orientation of unaggregated Methyl Orange at the rigid membrane surface.

In the case of the $2C_{16}N^+2C_1$ membrane matrix, the large blue shift observed is produced by dye aggregation, as also shown in Fig. 3. The λ_{\max} shift due to dye aggregation is small at temperatures above T_c , but is much larger at temperatures below T_c .

It is noteworthy that the two types of the rigid membrane matrix promote dye

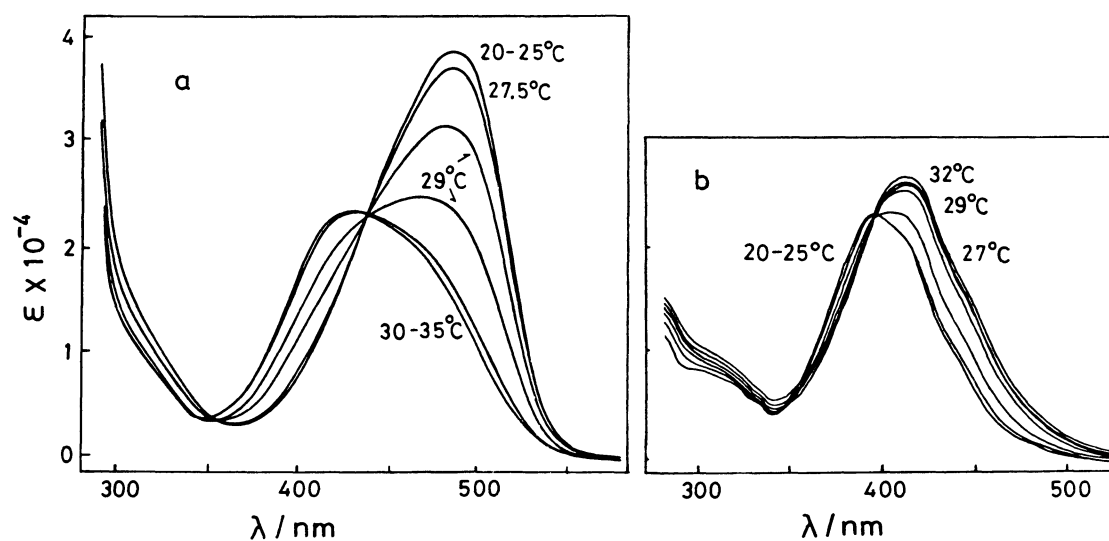


Fig. 1. Temperature dependence of absorption spectrum of Methyl Orange (2.5×10^{-5} M) bound to the bilayer membrane (2.5×10^{-4} M).

a. $L-1$ ($n = 4$), b. $2C_{16}N^+2C_1$.

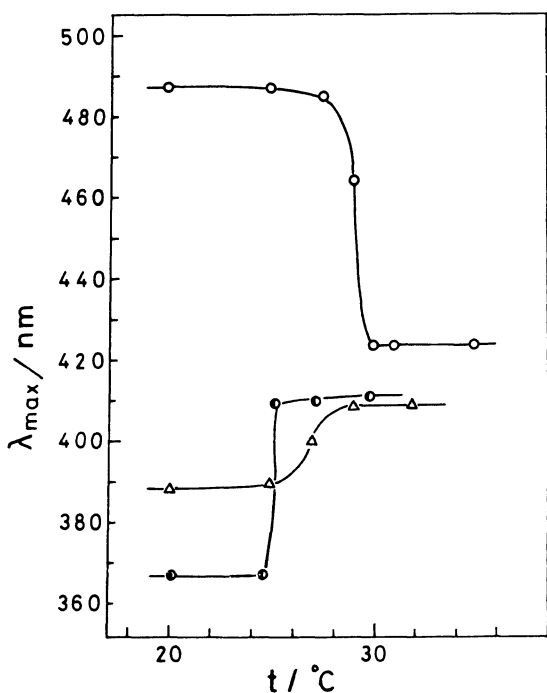


Fig. 2. Temperature dependence of λ_{max} of Methyl Orange (2.5×10^{-5} M) bound to the bilayer membrane (2.5×10^{-4} M).

○: $L-1$ ($n = 4$),
●: $L-1$ ($n = 2$),
△: $2C_{16}N^+2C_1$.

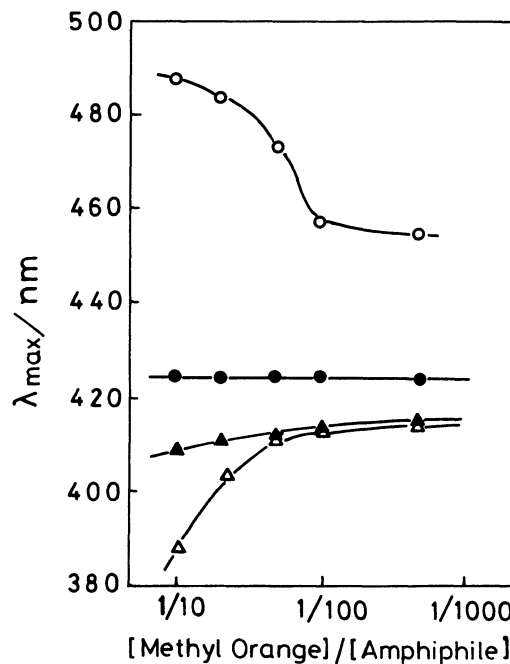
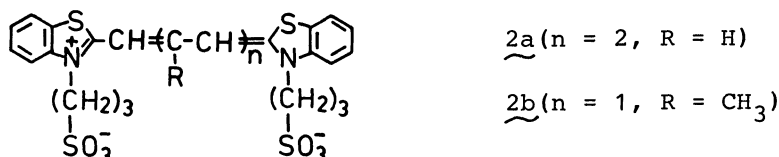


Fig. 3. Plots of λ_{max} of absorption spectra vs. $[Methyl\ Orange]/[amphiphile]$, Methyl Orange, 1.0×10^{-5} M (constant)

○: $L-1$ ($n = 4$) (20°C)
●: " (35°C)
△: $2C_{16}N^+2C_1$ (20°C)
▲: " (35°C)

aggregation in different orientations.

Spectral control by the bilayer membrane can be extended to a variety of cyanine and merocyanine dyes. For example, a considerable red shift was found for 2a bound to the membrane matrix of 1 ($n = 4$): $\lambda_{\max} = 721$ and 666 nm at temperature below and above T_c , respectively. More interestingly, drastic fluorescence enhancement (quantum yield, 0.64) has been achieved by the interaction of 2b with the bilayer membrane of 1 ($n = 4$; counterion, Cl^-). This value is 30 times larger than that observed in the CTAC micellar system and 250 times larger than those in methanol or in H_2O .¹¹⁾



In conclusion, the present study establishes that extensive and specific spectral control of Methyl Orange is possible. The λ_{\max} shift is presumably produced by the specific orientation and aggregation of the dye molecule at the membrane surface. This conclusion should pave a way to spectral control of dye molecules in general, as briefly described for cyanine dyes. These results should be useful not only in the model study¹⁴⁻¹⁶⁾ of biological chromophores such as membrane-bound chlorophyll but also from the practical point of view.

The authors are grateful to Miss Reiko Ando for her capable technical assistance.

References

- 1) Contribution No. 617 from Department of Organic Synthesis.
- 2) I. M. Klotz, G. P. Royer, and A. R. Sloniewsky, *Biochemistry*, 12, 4752 (1969).
- 3) T. Takagishi, Y. Nakata, and N. Kuroki, *J. Polym. Sci., Polym. Chem. Ed.*, 12, 807 (1974).
- 4) R. L. Reeves and S. A. Harkaway, "Micellization, Solubilization and Micro-emulsions", ed. by K. L. Mittal, Vol. 2, Plenum Press, New York (1977), p. 819.
- 5) The preparation of L-1 ($n = 4$) is described in Ref. 6. The other members of 1 were prepared by the same procedure.
- 6) T. Kunitake, N. Nakashima, M. Shimomura, Y. Okahata, K. Kano, and T. Ogawa, *J. Am. Chem. Soc.*, 102, 6642 (1980).
- 7) T. Kunitake and Y. Okahata, *J. Am. Chem. Soc.*, 99, 3860 (1977) and the subsequent papers.
- 8) T. Kunitake and S. Shinkai, *Adv. Phys. Org. Chem.*, 17, 435 (1980).
- 9) a) E. E. Jelly, *Nature*, 138, 1009 (1936). b) G. Scheibe, *Z. Angew. Chem.*, 50, 518 (1937).
- 10) A. H. Helz, *Adv. Colloid Interface Sci.*, 8, 237 (1977).
- 11) It has been reported that the fluorescence intensity of certain cyanine dyes in the micellar¹²⁾ and liposome¹³⁾ systems is 2 - 15 times greater than those in methanol or in water.
- 12) R. H-Baker, M. Grätzel, R. Steiger, *J. Am. Chem. Soc.*, 102, 847 (1980).
- 13) K. Onuki, K. Kurihara, Y. Toyoshima, M. Sukigara, *Bull. Chem. Soc. Jpn.*, 53, 1914 (1980).
- 14) A. G. Lee, *Biochem.*, 14, 4397 (1975).
- 15) K. Kurihara, Y. Toyoshima, and M. Sukigara, *J. Phys. Chem.*, 81, 1833 (1977).
- 16) A. Warshel, *J. Am. Chem. Soc.*, 101, 744 (1979)

(Received July 31, 1981)